

Polymorphisms in the nuclear factor kappa B gene association with recurrent embryo implantation failure

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ABSTRACT. Despite more than a century of intensive study, the mechanisms of successful pregnancy remain unclear. Recent research suggests that NF- κ B (nuclear factor kappa B) plays an important role in embryo implantation. In the current study, we aimed to identify SNPs that contribute to genetic susceptibility for recurrent implantation failure (RIF). Thus, we examined the potential associations between RIF and ten SNPs (rs28362491, rs3774932, rs1598856, rs230528, rs230521, rs3774956, rs4648055, rs3774964, rs4648068, and rs3774968) of the NF- κ B gene. Participants included 209 patients with RIF and 395 controls. Our results revealed that there were statistically significant differences observed in the allelic and genotypic frequencies of the

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rs28362491 promoter in the NF-κB gene. The frequency of the del/ del genotype was significantly higher in RIF patients than in healthy controls (P = 0.004). Compared with healthy controls, the RIF patients carried a higher frequency of the rs28362491 del allele (P = 0.010). Furthermore, strong linkage disequilibrium was observed in the three identified haplotype blocks (D' > 0.9). Particularly, in block 1 (rs230528-rs230521), the A-C haplotype occurred significantly more frequently (P = 0.029) in subjects with RIF (P = 0.0003). In contrast, the A-G haplotype occurred significantly less frequently (P = 0.008) in RIF subjects. These findings support an important role for G-712A polymorphisms of NF-κB in RIF, and may guide future studies that aim to characterize genetic risk factors for RIF.

Key words: Repeated implantation failure; Nuclear factor kappa B; Single nucleotide polymorphism; SNP

INTRODUCTION

Recurrent implantation failure (RIF) refers to the failure to achieve a clinical pregnancy after transfer of at least four good-quality embryos from a minimum of three fresh or frozen cycles in a woman under the age of 40 years (Coughlan et al., 2014). Implantation failure is related to either maternal factors or embryonic causes. Maternal factors include uterine anatomic abnormalities, thrombophilia, non-receptive endometrium, and immunological factors. Failures of implantation due to embryonic causes are associated with either genetic abnormalities or other factors intrinsic to the embryo that impair their ability to develop in utero, hatch, and implant (Simon and Laufer, 2012).

Increased incidence of chromosomal translocations, mosaics, inversions, and deletions of genetic material have been reported in patients with RIF (Raziel et al., 2002). Moreover, almost twice as many chromosomal abnormalities were detected in embryos from RIF patients than in embryos of controls (67.4% versus 36.3%) (Pehlivan et al., 2003), and significantly more complex chromosomal abnormalities that involve three or more chromosomes were reported in cleaved embryos of RIF patients than in those of controls (Voullaire et al., 2007).

Nuclear factor kappa B (NF- κ B) proteins are a family of structurally related eukaryotic transcription factors that are persistently active in the pathogenesis of numerous malignancies (Lind et al., 2001; Rueda et al., 2006; Hua et al., 2014) in addition to RIF (Cakmak et al., 2009; Dharmaraj et al., 2010). The NF- κ B family consists of five members in mammals that include the following: NF-KB1 encoding for p50, NF-KB2 encoding for p52, RELA encoding for p65, REL encoding for c-Rel, and RELB encoding for Rel-B. The most common dimer formed by these members is the p65/p50 heterodimer (Lu et al., 2012). The human NF-KB1 gene is located at chromosome region 4q24. Several researchers have shown a relationship between gastric cancer and polymorphisms in NF-KB1 promoter regions (Inagaki et al., 2009; Tchernitsa et al., 2010; Hua et al., 2014). Therefore, the aim of the current study was to investigate the possible associations between NF-KB1 single nucleotide polymorphisms (SNPs) and susceptibility to RIF.

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MATERIAL AND METHODS

Subjects

The study group consisted of 209 women diagnosed with RIF from March 2010 to June 2014 on the basis of the following criteria: all women had \geq 3 failed embryo transfers (ET) (ranging from 3-6) with transfers of \geq 1 high-quality (grades I and II) fresh or frozen-thawed embryos each; and all patients underwent at least two fresh ET. Embryos were classified as we previously reported as follows: grade 1 = perfectly symmetrical with no fragmentation; grade 2 = perfectly symmetrical with slight fragmentation (<20%) fragmentation of the total embryonic volume); grade 3 = uneven blastomeres with no fragmentation; and grade 4 = uneven blastomeres with gross fragmentation (>20%) fragments) (Creus et al., 2003). Embryos of grades 1 or 2 were considered of high quality and were at the 4-cell or 8-cell stage on Day 2 or 3 post-fertilization, respectively. For purposes of statistical comparison and to objectively quantify embryo quality, embryos of grades 1-4 were scored 2.5, 2, 1.5 and 1, respectively, as previously reported (Puerto et al., 2003). For the final analysis, the embryo scores per ET were assessed as the mean values of the scores given to each of the transferred embryos. The IVF group (N = 396) consisted of patients undergoing a first IVF-ET cycle before and after each ET, and their success was used to define patients as RIF. This study was approved by the Ethical Committee of the Fourth Hospital of Xi'an (Xi'an, China), and written informed consent was obtained from all participants.

Genotyping

Whole peripheral blood was obtained from each study participant, and genomic DNA was extracted from blood leukocytes using the E.Z.N.A. Blood DNA Midi Kit (Omega BioTek, Norcross, GA, USA) according to manufacturer protocol. We then searched for SNPs with a minor allele frequency (MAF) >0.05 in three different linkage disequilibrium (LD) blocks. Genotyping was carried out for all SNPs using the MassARRAY platform (Sequenom, San Diego, CA, USA). Specifically, SNPs were genotyped using high-throughput matrix-assisted laser desorption ionization time-of-flight (MALDI-TOF) mass spectrometry. Next, the resulting spectra were processed with Typer Analyzer software (Sequenom), and genotype data were generated for each sample. The final genotype call rate of each SNP was >98%, and the overall genotype call rate was 98.6%, ensuring the reliability of further statistical analysis.

Statistical analysis

All statistical analyses were carried out using SPSS 16.0 software (SPSS Inc., Chicago, IL, USA). Hardy-Weinberg equilibrium (HWE) was evaluated, and associations between cases and controls for each polymorphism were assessed by Pearson's chi-square tests. A Bonferroni correction was used for multiple tests, wherein the P values were divided by the total number of loci or haplotypes. Pairwise LD statistics (D' and R²) and haplotype frequencies were computed using Haploview 4.0 (http://www.broad.mit.edu/mpg/haploview) to construct haplotype blocks.

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RESULTS

No significant deviations from HWE were found between cases and controls for any of the SNPs identified herein. LD analyses of the patients and controls revealed that seven downstream SNPs were located in a different haplotype block (Fig. 1). Furthermore, LD analysis data showed that rs230528 and rs230521 were located in block 1; rs3774956 and rs4648055 were located in block 2; and that rs4648068, rs3774964, and rs3774968 were located in block 3. The genotype distribution, allelic frequencies, and haplotypes of the RIF patients and healthy controls are shown in Tables 1-4.



Figure 1. Linkage disequilibrium (LD) plot of the ten single nucleotide polymorphisms (SNPs) in the NF κ B gene.

Variable	Group	Genotype [N (%)]			Allele [N (%)]		Pa	Pb
		ins/ins	ins/del	del/del	ins	del		
rs28362491	Control	106 (26.84)	214 (54.18)	75 (18.99)	426 (53.92)	364 (46.08)	0.004	0.010
	Case	49 (23.44)	95 (45.45)	65 (31.10)	193 (46.17)	225 (53.83)		
		AA	AG	GG	A	G		
rs3774932	Control	151 (38.23)	188 (47.59)	56 (14.18)	490 (62.03)	300 (37.97)	0.831	0.555
	Case	76 (36.36)	100 (47.85)	33 (15.79)	252 (60.29)	166 (39.71)		
		CC	CT	TT	С	Т		
rs1598856	Control	115 (29.26)	178 (45.29)	100 (25.45)	408 (51.91)	378 (48.09)	0.293	0.755
	Case	52 (25.00)	108 (51.92)	48 (23.08)	212 (50.96)	204 (49.04)		
		AA	AC	CC	A	С		
rs230528	Control	89 (22.53)	189 (47.85)	117 (29.62)	367 (46.46)	423 (53.54)	0.498	0.485
	Case	47 (22.49)	109 (52.15)	53 (25.36)	203 (48.56)	215 (51.44)		
		CC	CG	GG	С	G		
rs230521	Control	148 (37.47)	179 (45.32)	68 (17.22)	475 (60.13)	315 (39.87)	0.001	0.0002
	Case	106 (50.72)	85 (40.67)	18 (8.61)	297 (71.05)	121 (28.95)		
		CC	CT	TT	С	Т		
rs3774956	Control	46 (11.65)	177 (44.81)	172 (43.54)	269 (34.05)	521 (65.95)	0.993	0.956
	Case	25 (11.96)	93 (44.50)	91 (43.54)	143 (34.21)	275 (65.79)		
		AA	AG	GG	A	G		
rs4648055	Control	115 (29.11)	178 (45.06)	102 (25.82)	408 (51.65)	382 (48.35)	0.887	0.759
	Case	57 (27.27)	98 (46.89)	54 (25.84)	212 (50.72)	206 (49.28)		
		AA	AG	GG	A	G		
rs3774964	Control	112 (28.35)	210(53.16)	73 (18.48)	434 (54.94)	356 (45.06)	0.137	0.773
	Case	66 (31.58)	94 (44.98)	49 (23.44)	226 (54.07)	192 (45.93)		
		AA	AG	GG	A	G		
rs4648068	Control	106 (26.84)	214 (54.18)	75 (18.99)	426 (53.92)	364 (46.08)	0.004	0.010
	Case	49 (23.44)	95 (45.45)	65 (31.10)	193 (46.17)	225 (53.82)]	
		AA	AG	GG	A	G]	
rs3774968	Control	100 (25.32)	214 (54.18)	81 (20.51)	414 (52.41)	376 (47.59)	0.119	0.637
	Case	65 (31.10)	95 (45.45)	49 (23.44)	225 (53.83)	193 (46.17)		

^aP values for genotype frequency differences between RIF patients and controls. ^bP values for allele frequency differences between RIF patients and controls. Alpha values were adjusted by Bonferroni correction and statistically significant results are shown as P < 0.005.

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Table 2. NF- κ B haplotype in block 1 frequencies and the results of their associations with risk of RIF.								
Variable	Case (209)		Contro	d (395)	P value ^a	OR, 95%CI		
	N	%	N	%				
C-C	106	50.72	208	52.66	0.650	0.925, 0.662-1.294		
A-G	59	28.22	154	38.99	0.008	0.616, 0.428-0.885		
A-C	42	20.10	29	7.34	0.0003	2.498, 1.500-4.160		

^aBased on comparisons of frequency distributions of all haplotypes for the combination of SNPs.

Table 3. NF- κ B haplotype in block 2 frequencies and the results of their associations with risk of RIF.

Variable	Case (209)		Contro	l (395)	P value ^a	OR, 95%CI
	N	%	N	%		
T-A	105	50.24	204	51.65	0.742	0.945, 0.676-1.322
C-G	71	33.97	135	34.18	0.959	0.991, 0.696-1.411
T-G	32	15.31	56	14.18	0.479	0.843, 0.524-1.354

^aBased on comparisons of frequency distributions of all haplotypes for the combination of SNPs.

Table 4. NF-κB haplotype in block 3 frequencies and the results of their associations with risk of RIF.								
Case (209)		Contro	ol (395)	P value ^a	OR, 95%CI			
Ν	%	N	%					
111	53.11	181	45.82	0.088	1.339, 0.957-1.874			
95	45.45	177	44.81	0.880	1.026, 0.733-1.437			
	IF-κB haplotype Case N 111 95	N % 111 53.11 95 45.45	Case (209) Control N % N 111 53.11 181 95 45.45 177	N % N % 111 53.11 181 45.82 95 45.45 177 44.81	N % N % 111 53.11 181 45.82 0.088 95 45.45 177 44.81 0.880			

^aBased on comparisons of frequency distributions of all haplotypes for the combination of SNPs.

These results demonstrated significant differences in the genotype and allele frequency distributions of rs28362491 between the RIF patients and healthy controls. Particularly, the frequency of the del/del genotype was significantly higher in RIF patients than that in healthy controls (P = 0.004). Specifically, compared with healthy controls, the RIF patients carried a higher frequency of the rs28362491 del allele (P = 0.010) (Table 1).

The differences in the distributions of genotype frequencies of the haplotypes (block 1) between RIF subjects and healthy controls were significant. Specifically, the A-C (rs230528-rs230521) haplotype occurred significantly more frequently in subjects with RIF than in healthy controls (P = 0.0003). Additionally, the A-G haplotypes occurred significantly less frequently in RIF subjects than in healthy con (P = 0.008).

DISCUSSION

One of the key goals in linkage disequilibrium and association studies is to identify common risk variants in different populations. To determine if common risk variants exist for RIF patients in distinct populations, we compared our results to those of previous studies. The NF- κ B pathway is a nearly ubiquitous pathway responsible for mediating DNA transcription, and thereby cell function. Additionally, NF- κ B is a major transcriptional regulator of immune responses, apoptosis, and cell growth control genes. In previous studies, evidence has indicated that NF- κ B plays a pivotal role in the development of RIF (Cakmak et al., 2009; Dharmaraj et al., 2010). Importantly, our results herein provide direct evidence that a genetic change in NF- κ B is linked to RIF in humans.

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Genetic variation contributes to different disease susceptibility. The functional significance of several NF-kB1 SNPs, particularly rs28362491 (-94ins/del ATTG), has been well elucidated (Hogan, 1988). The NF-κB1 -94ins/del ATTG promoter polymorphism is an insertion/deletion of four bases in the promoter region of the NF-KB gene that encodes for the p50 subunit, and the variant allele containing this deletion produces lower transcript levels of the p50 subunit (Karban et al., 2004). The NF-κB1 rs28362491 promoter polymorphism is the only known functional polymorphism in the NF-KB family (van der Heiden et al., 2010; Vogel et al., 2011), and this polymorphism has been shown to be associated with both inflammation and coronary syndrome (Tjønneland et al., 2007). In the current study, we found a significant difference in the genotype distribution of rs28362491 between those with RIF and controls. Specifically, subjects with RIF had a significantly higher frequency of the rs28362491 del allele than that of controls. Based on these observations, our data indicate that the NF- κ B rs28362491 genotype is associated with RIF. To our knowledge, this is the first study to identify a significant association between rs28362491 in the promoter region of the NF-κB gene and RIF. However, future in vivo investigations will be required to precisely elucidate the mechanism by which the functional SNPs of NF-kB affect protein function and levels.

In the haplotype analysis, a significant association between NF- κ B gene haplotypes (rs230528-rs230521) and RIF was found. Specifically, the A-C (rs230528-rs230521) haplotype occurred significantly more frequently in subjects with RIF than in controls. These results indicate that people with the A-C haplotype are more prone to RIF. Additionally, the A-G haplotypes occurred significantly less frequently in RIF subjects. These results indicate that patients with the A-G haplotype of the NF- κ B gene were less prone to RIF, which further suggests that this haplotype may have protective effects against RIF. However, the functional effects of the different NF- κ B haplotypes remain unknown, though they may play an important role in the development of RIF. Evidence from the current study supports the association of NF- κ B polymorphisms and RIF.

In the current study, we found that in a large and homogeneous population, NF- κ B gene polymorphisms (rs28362491) were associated with RIF. These findings encourage future RIF investigations into the functional polymorphisms of NF- κ B within in larger sample populations.

Conflicts of interest

The authors declare no conflicts of interest.

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