

Molecular cloning and expression analysis of a novel BCCP subunit gene from *Aleurites moluccana*

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ABSTRACT. Aleurites moluccana L. is grown as a roadside tree in southern China and the oil content of its seed is higher than other oil plants, such as Jatropha curcas and Camellia oleifera. A. moluccana is considered a promising energy plant because its seed oil could be used to produce biodiesel and bio-jet fuel. In addition, the bark, leaves, and kernels of A. moluccana have various medical and commercial uses. Here, a novel gene coding the biotin carboxyl carrier protein subunit (BCCP) was cloned from A. moluccana L. using the homology cloning method combined with rapid amplification of cDNA end (RACE) technology. The isolated full-length cDNA sequence (designated AMaccB) was 1188 bp, containing a 795-bp open reading frame coding for 265 amino acids. The deduced amino acid sequence of AM-accB contained a biotinylated domain located between amino acids 190 and 263. A. moluccana BCCP shows high identity at the amino acid level to its homologues in other higher plants, such as Vernicia fordii, J. curcas, and Ricinus communis (86, 77, and 70%, respectively), which all contain conserved domains for ACCase activity. The expression of the AM-accB gene during the middle stage of development and maturation in A. moluccana seeds was higher than that in early and later stages.

Genetics and Molecular Research 14 (3): 9922-9931 (2015)

Analysis of a new BCCP subunit gene from Aleurites moluccana

The expression pattern of the *AM*-accB gene is very similar to that of the oil accumulation rate.

Key words: *Aleurites moluccana* L.; BCCP subunit gene; Molecular cloning; Sequence analysis; Expression analysis

INTRODUCTION

Aleurites moluccana L. is a perennial, woody oil plant belonging to the family Euphorbiaceae and it is widely distributed in the tropics and subtropics worldwide. The oil content of its seeds is 66.22% (Liu et al., 2008), which is higher than other oil plants, such as peanut (Guo et al., 2012), rapeseed (Zhang et al., 2007), and *Camellia oleifera* (Wang et al., 2008). *A. moluccana* is a promising energy plant because of its high oil content, which could be used for biodiesel and bio-jet fuel production.

The oil content of plant seeds is affected by both genotype and environmental conditions, with the genotype playing a leading role. In the process of oil biosynthesis, there are a series of related pathways and catalysis of various enzymes. There are two distinct subcellular forms of acetyl-CoA carboxylase (ACCase) in plant cells; one is homomeric ACCase, mainly located in the cytosol and expressed constitutively, and the other heteromeric ACCase, is found in plastids and is expressed in developing seeds (Thelen et al., 2000; 2001; Sasaki and Nagano, 2004). The production of malonyl-CoA catalyzed by ACCase is the rate-limiting step in the process of fatty acid biosynthesis (Thelen et al., 2000; Fukuda et al., 2013). Previous studies have confirmed that heteromeric ACCase plays an important regulatory function during oil synthesis and accumulation in plant seeds (Thelen et al., 2000; Dong et al., 2004; Hu et al., 2009; Liu et al., 2009; Li et al., 2011). Homomeric ACCase is composed of a large multifunctional polypeptide (Sasaki and Nagano, 2004) and heteromeric ACCase is composed of a biotin carboxyl carrier protein (BCCP), biotin carboxylase (BC), and carboxyltransferase (CT), which consists of α - and β -subunits. Among the subunits of ACCase, BCCP subunit serves as the donor of activated carboxyl group and shuttles the tethered biotin between BC and CT domains, which is important for *de novo* fatty acid biosynthesis (Ke et al., 2000; Cronan, 2002; Sasaki and Nagano, 2004).

To date, the genes of many plant ACCase subunits have been cloned and analyzed, with the full-length cDNAs of ACCase subunit genes being in the range 1000 to 2947 bp and showing 50 to 98.8% sequence identity with each other in the same gene of different plants (Tan et al., 2010; Gu et al., 2011). The synthesis and accumulation of oil in plant seeds is closely related to the expression level of ACCase genes during seed development (Chen et al., 2009; Rogalski and Carrer, 2011).

In many plants, such as *Arabidopsis*, *Brassica napus*, *Vernicia fordii*, *Jatropha*, soybean, and castor bean, the BCCP subunit gene of ACCase has been successfully cloned (Choi et al., 1995; Elborough et al., 1996; Reverdatto et al., 1999; Thelen et al., 2001; Gu et al., 2011; Cui et al., 2012). The expression patterns of the BCCP subunit gene vary greatly in different tissues and organs of plants. *Arabidopsis* expresses two paralogs of the BCCP subunit, where BCCP 1 is coded by *BCCP1* and BCCP 2 is coded by *BCCP2*. The *BCCP1* transcript is expressed constitutively, while *BCCP2* is predominant in flowers and siliques and its overall abundance is approximately 4-fold lower than that of *BCCP1* (Thelen et al., 2000). The development of the endosperm and embryo is disrupted when *BCCP1* is knocked-out during *Arabidopsis* seed development, and fatty acid accumulation decreases and normal vegetative

Genetics and Molecular Research 14 (3): 9922-9931 (2015)

w. I. Muan et al.	W.Y	Xuan	et	al.	
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plant growth is severely affected when BCCP1 accumulation is reduced at the protein level (Li et al., 2011). These results indicate that BCCP genes are involved in de novo lipid synthesis storage and accumulation (Elborough et al., 1996; Thelen et al., 2000; Gu et al., 2011). The lipid content of *A. moluccana* seeds is higher than that of many other plants, but little is known about the sequence structure of the BCCP subunit gene and its expression profile during seed development and oil accumulation in *A. moluccana*.

In this study, in order to explore the role of the BCCP subunit gene during seed development and oil accumulation in *A. moluccana*, the full-length cDNA sequence for the BCCP subunit was obtained successfully by employing homology cloning and the rapid amplification of cDNA end (RACE) technique. The expression profile of the gene and the oil content of the seeds were determined using quantitative real-time PCR (qRT-PCR) and Soxhlet, respectively. The results provide a basis for further research about nucleotide polymorphisms of the BCCP gene and molecular markers linked to the oil content of *A. moluccana* seeds.

MATERIAL AND METHODS

Plant material

Young leaves and seeds at various stages of development were collected from *A. mo-luccana L.* plants distributed in Guangxi University and stored at -80°C until required.

Total RNA extraction and cDNA synthesis

Total RNA was extracted from young leaves and cDNA was synthesized using the RevertAidTM First Strand cDNA Synthesis kit (Fermentas, Lithuania) with 5 μ g of RNA as a template, and following the manufacturer protocol.

Degenerate primer design

A pair of degenerate primers was designed using the online software CODEHOP (http://blocks.fhcrc.org/blocks/codehop.html) according to the conserved amino acid sequences of BCCP in related species. The primer sequences are described in Table 1.

cDNA of the gene coding for the BCCP subunit cloned from A. moluccana L.

The middle fragment of cDNA coding for the BCCP subunit was amplified by PCR using the degenerate primer pair described in Table 1. PCR products were cloned into a pUCm-T vector (Sangon, Shanghai, China), and transferred into competent cells of *Escherichia coli* strain DH51 α . The positive clones were screened out using blue-white selection, confirmed by PCR, and sequenced. The sequence was submitted to GenBank to identify similar genes through Blastx. To obtain the full-length cDNA sequence, a set of gene specific primers were designed according to the sequence of the middle fragment of cDNA for 5'- and 3'-RACE. These primers are shown in Table 1. The amplified sequences were cloned, sequenced, and assembled. Synthesis of all primers and sequencing were conducted by the Sangon.

Genetics and Molecular Research 14 (3): 9922-9931 (2015)

Objective gene sequence analysis

The location of the open reading frame (ORF) and DNA translation were predicted by using the online software ExPASy (http://web.expasy.org/translate/) and DNAMAN (Lynnon Biosoft, USA). Amino acid sequences of homologs from different plants were obtained from GenBank (http://www.ncbi.nlm.nih.gov/) and aligned using Biology Workbench 3.2 (http:// workbench.sdsc.edu/AlignmentTools). The software ExPASy (www.expasy.org) was used to analyze the theoretical isoelectric point (pI) and molecular weight of the predicted protein. The online software ProtScale (http://web.expasy.org/ProtScale/) and Predict Protein (http:// www.PredictProtein.org/) were used to analyze the hydropathicity and secondary structure composition of the predicted protein.

Real-time RT-PCR analysis of gene expression

Total RNA was isolated from the *A. moluccana* endosperm using RNAprep Pure Plant kits (TIANGEN Biotechnology Company, Beijing, China) according to the manufacturer protocol. First strand cDNA was synthesized using the RevertAid-TM First Strand cDNA synthesis kit (Fermentas) according to the manufacturer protocol. qRT-PCR was performed to examine expression of the BCCP subunit gene. The 18SrRNA gene was used as an internal control in *A. moluccana*. The specific primers for the internal control gene and the target gene used for qRT-PCR are shown in Table 1. The amplified fragment lengths of the control and target gene were 110 bp and 217 bp, respectively. Finally, cDNA was amplified using real-time PCR. All amplifications were performed using SYBR Premix ExTaq II (TaKaRa Biotechnology Company, Dalian, China). SYBR Green II assay reactions were performed in a 10 μ L volume and each reaction contained 5 μ L SYBR Premix ExTaq II, 400 nM concentration of each primer, and cDNA. PCRs were performed with four biological replicates. Gene expression levels were evaluated using the delta/delta calculation method (Livak and Schmittgen, 2001). The data were then processed using the base 10 logarithm and Excel mapping.

Seed oil content by Soxhlet analysis

Oil content was quantified using Soxhlet analysis. *A. moluccana* seed samples were harvested at various stages of seed development and shelled and dried at 80°C for use to determine the oil content. Seed oil content was measured, and the mass of each measurement was converted to a percentage of the total weight of each sample. All samples were measured in triplicate (3 seeds for each replication) and the results averaged.

RESULTS

Full length cDNA cloning of the BCCP subunit gene

A middle fragment of the BCCP subunit gene (approximately 500 bp in length; Figure 1A) and its 5'-cDNA end (approximately 400 bp; Figure 1B) and a 3'-cDNA end (approximately 450 bp; Figure 1C) were amplified using RT-PCR with total RNA isolated from young leaves as a template and the degenerate, 5'-RACE, and 3'-RACE primers, respectively (Table 1). Sequencing results showed that the lengths of these fragments are 456, 473, and 494 bp,

Genetics and Molecular Research 14 (3): 9922-9931 (2015)

respectively. A full-length cDNA sequence of 1188 bp was obtained by assembling the fragments and was termed *AM*-accB. *AM*-accB contained a 795-bp ORF coding for 265 amino acids, a 5'-UTR of 202 bp, and a 3'-UTR of 191 bp.

 Table 1. Primers* used for PCR.

Name	Sequence
Degenerate primers P1	5'-TACTAAGATGCAAGCTCAAYTNAAYGARGT-3'
Degenerate primers P2	5'-CTATAAAAAGTTCCAGCCATNGGRCAYTT-3'
5' RACE GSP1	5'-TTTGCTCCTCCAAGTTCCAC-3'
5' RACE GSP2	5'-CAAGTGTCAGACCTTGTCAAGC-3'
5' RACE GSP3	5'-TTCTGTCCCTGTGCTAGACAAC-3'
3' RACE GSP1	5'-GTCAACTAAACGGCATTCCC-3'
3' RACE GSP2	5'-CTTATTTTAGTGTGTGCTTCCG-3'
18srRNA primers P1	5'-TGTTGCTTTTAGGACTCCGCCGG-3'
18srRNA primers P2	5'-GTGGTGCCCTTCCGTCAATTCCT-3'
Q-RT-PCR P1	5'-TCCAGAGTTCCATCATCCGAGGT-3'
Q-RT-PCR P2	5'-GCGCTGCAATGACAGGAGGT-3'
*Y = C/T, N = A/C/G/T.	
M 1 2 3	1234 M M 123
600 bp -	- 700 bp - 500 bp 450 bp -

Figure 1. PCR amplification of the BCCP subunit in *Aleurites moluccana* **A.** *Lanes 1* to 3: products of the middle fragment of the BCCP subunit sequence; **B.** *lanes 1* to 4: 5'-cDNA products; **C.** *lanes 1* to 3: 3'-cDNA products; *lane M*: DNA marker.

В

Sequence analysis of the BCCP subunit gene

A

Homology analysis by Blastx revealed that *AM*-accB codes for the BCCP subunit of *A. moluccana* and shares high identity with sequences from *V. fordii, J. curcas,* and *R. communis,* (86, 77, and 70%, respectively), based on amino acid sequence comparison. The BCCP subunit in *A. moluccana* (AM-BCCP) is a chloroplast located protein and its deduced amino acid sequence contains a biotinylated domain EAMKLMN located at the C-terminus between amino acids 190 and 263 (Figure 2). It has a molecular weight and theoretical isoelectric point of 28.2 kDa and 7.61, respectively. The predicted hydropathicity of AM-BCCP showed a minimum of -2.667 and a maximum of 1.844. When the window size (using linear weight variation model) was 11, it had two peaks that scored >1.5 and 13 peaks that scored <0 (Figure 3). The results indicate that AM-BCCP has water-soluble properties. Analysis using the online software Predict Protein indicated that AM-BCCP secondary structure is composed of 12.08% helix, 17.74% sheet, and 70.19% loops, with one helix, seven sheets, and nineteen loops with a confidence value >5, and it has no potential transmembrane domain. Analysis using the online software SignalP (4.1 Server) showed that the AM-BCCP protein has no signal peptides.

С

Genetics and Molecular Research 14 (3): 9922-9931 (2015)

AM-BCCP	MASISVPCPNTASVARFGS	50
Vernicia fordii	MASISVPCPKTALVARFGSNATNPLPKSTISFPPRASSLLGFQWSNRKQS	
Jatropha curcas	MASISVPCPKTILVARIGSNVENPLPNSTVSFPPLTSNLLGFQWCNWKQS	
Ricinus communis	MASTSVPCPKTSFIARIGSNASNSLPKHTISFPSRGFKWPIRKQS	
	*** *****:*: :**:** *::*: *:**:: **:*	
AM-BCCP	PVTKMQAQLNEVLAEKSSNSVPVLDNNSRVPSSEVKDEPAESNTPDASAI	100
Vernicia fordii	PVTKVQAQLNEVLAEKSSNSVPVLDNNSKVPSSEQKDEPAENNIPDASAI	
Jatropha curcas	RVTKMQAKLNEVLLEKSSNSVPVLETNSNVPTPDEKDESTESNILDASQI	
Ricinus communis	AVTKVKAQLNEVIVKGSSNSVPMLDTNSKLAPSEVKDEAVVKNVPDASAI	
	::*:*:::: ******:*::**.::::: ***:* ***	
AM-BCCP	SAFMTQVSDLVKLVDSRDITELQLKQLGCELIIRKKEALQQPLP-PVIA	150
Vernicia fordii	SAFMTQVSDLVKLVDSRDITELQLKQLGCELIIRKKEALQQPPP-PVIA	
Jatropha curcas	SAFMTQVSDLVKLVDSRDITELQLKQLGCQLIIRKKEALQQPPPAAPVIA	
Ricinus communis	TEFMTQVSDLVKLVDSRDITELQLKQLDCEIIIRKKEALQ-PPPAAQVIS	
	: ************************************	
	100	
	150	
AM-BCCP	AQPSYQHALLPSPPPAAPAFAPPSSTSPP-APALPSPAKTNTSRTP-LKC	200
AM-BCCP Vernicia fordii	AQPSYQHALLPSPPPAAPAFAPPSSTSPP-APALPSPAKTNTSRTP-LKC AQPSYQHAMLPSPPLAAPASAPPSSTPPPAPALPSPAKSSTSSHPQLKC	200
AM-BCCP Vernicia fordii Jatropha curcas	AQPSYQHALLPSPPPAAPAFAPPSSTSPP-APALPSPAKTNTSRTP-LKC AQPSYQHAMLPSPPLAAPASAPPSSTPPPPAPALPSPAKSSTSSHPQLKC MPPSYQHTMLPSPPPAAPASAPPSS-PPLALALPPPAKTSSSSHPPLKC	200
AM-BCCP Vernicia fordii Jatropha curcas Ricinus communis	AQP SYQHALLP SPPPAAP AF APP SSTSPP-APALP SPAKTNTSRTP-LKC AQP SYQHAMLP SPPLAAP AS APP SSTPPPPAPALP SPAKSSTSSHPQLKC MPP SYQHTMLP SPPPAAPASAPP SS-PPLALALPPPAKTSSSSHPPLKC MPP SYQHVMLPPPP SAAP-PPSS-AP-APALP SPAKAVTSSHPAFKC	200
AM-BCCP Vernicia fordii Jatropha curcas Ricinus communis	AQPSYQHALLPSPPPAAPAFAPPSSTSPP-APALPSPAKTNTSRTP-LKC AQPSYQHAMLPSPPLAAPASAPPSSTPPPPAPALPSPAKSSTSSHPQLKC MPPSYQHTMLPSPPPAAPASAPPSS—PPLALALPPPAKTSSSSHPPLKC MPPSYQHVMLPPPPSAAP—PPSS—AP-APALPSPAKAVTSSHPAFKC *****.:**.** *** *** *** .* ****.***: :* * :**	200
AM-BCCP Vernicia fordii Jatropha curcas Ricinus communis	AQP SYQHALLP SPPPAAP AF APP SSTSPP-APALP SPAKTNTSRTP-LKC AQP SYQHAMLP SPPLAAP AS APP SSTPPPPAPALP SPAKSSTSSHPQLKC MPP SYQHTMLP SPPPAAP AS APP SS-PPLALALPPPAKTSSSSHPPLKC MPP SYQHVMLPPPP SAAP-PPSS-AP-APALP SPAKAVTSSHPAFKC *****.:**.** *** *** *** .* * ***.***: :* * :**	200
AM-BCCP Vernicia fordii Jatropha curcas Ricinus communis AM-BCCP	AQPSYQHALLPSPPPAAPAFAPPSSTSPP-APALPSPAKTNTSRTP-LKC AQPSYQHAMLPSPPLAAPASAPPSSTPPPPAPALPSPAKSSTSSHPQLKC MPPSYQHTMLPSPPPAAPASAPPSS—PPLALALPPPAKTSSSSHPPLKC MPPSYQHVMLPPPPSAAP—PPSS—AP-APALPSPAKAVTSSHPAFKC *****.:**.** *** *** .*** .** ***.***: :* * :*** PMAGTFYRLPAPGEPPFVKVGDKVQKGQVVCIIEAMKLMNEIEADQSGTI	200
AM-BCCP Vernicia fordii Jatropha curcas Ricinus communis AM-BCCP Vernicia fordii	AQP SYQHALLP SPPPAAP AF AP AF ST SPP-AP ALP SPAKTNT SRTP-LKC AQP SYQHAMLP SPPLAAP AS APP SSTPPPPAP ALP SPAKS ST SSHPQLKC MPP SYQHTMLP SPPPAAP AS APP SS — PPLALALPPPAKT SSS SSHPPLKC MPP SYQHVMLPPPP SAAP — PPSS — AP-AP ALP SPAKAVT SSHPAFKC *****.:**.** *** *** .** *** .** *** ***	200 250
AM-BCCP Vernicia fordii Jatropha curcas Ricinus communis AM-BCCP Vernicia fordii Jatropha curcas	AQPSYQHALLPSPPPAAPAFAPPSSTSPP-APALPSPAKTNTSRTP-LKC AQPSYQHAMLPSPPLAAPASAPPSSTPPPPAPALPSPAKSSTSSHPQLKC MPPSYQHTMLPSPPPAAPASAPPSS-PPLALALPPPAKTSSSSHPPLKC MPPSYQHVMLPPPPSAAP-PPSS-AP-APALPSPAKAVTSSHPAFKC *****.:**.** *** **** .* ****.***: :* * :*** PMAGTFYRLPAPGEPPFVKVGDKVQKGQVVCIIEAMKLMNEIEADQSGTI PMAGTFYRCPAPGEPPFVKVGDKVQKGQVVCIIEAMKLMNEIEADQSGTI PMAGTFYRNPAPGEPPFVKVGDKVQKGQVVCIIEAMKLMNEIEADQSGTI	200 250
AM-BCCP Vernicia fordii Jatropha curcas Ricinus communis AM-BCCP Vernicia fordii Jatropha curcas Ricinus communis	AQP SYQHALLP SPPPAAP AF AP AF AP P S ST SPP-AP ALP SPAKTNT S RTP-LKC AQP SYQHAMLP SPPLAAP AS AP P S ST PPP AP ALP SPAK ST S SHPQLKC MPP SYQHTMLP SPPPAAP AS AP P S S-PPLALALP PPAKT SS S SHPPLKC MPP SYQHVMLPPPP SAAP-PPS S-AP-AP ALP SPAKAVT S SHPAFKC *****.:**.** *** *** .** *** .** *** .** *** PMAGTFYRLP APGEPPFVKVGDKVQKGQVVCIIE AMKLMNE IE ADQ SGT I PMAGTFYRCP APGEPPFVKVGDKVQKGQVVCIIE AMKLMNE IE ADQ SGT I PMAGTFYRNP APGEPPFVKVGDKVQKGQVVCIIE AMKLMNE IE ADQ SGT I PMAGTFYRCP APGEPPFVKVGDKVQKGQVVCIIE AMKLMNE IE ADQ SGT I PMAGTFYRCP APGEPPFVKVGDKVQKGQVVCIIE AMKLMNE IE ADQ SGT I	200
AM-BCCP Vernicia fordii Jatropha curcas Ricinus communis AM-BCCP Vernicia fordii Jatropha curcas Ricinus communis	AQP SYQHALLP SPPPAAP AF AP AF AP P S ST SPP – AP ALP SPAKTNT S RTP – LKC AQP SYQHAMLP SPPLAAP AS AP P S ST PPP AP ALP SPAK S ST S SH PQLKC MPP SYQHTMLP SPPPAAP AS AP P S S — AP – AP ALP SPAK AVT S SH PAFKC *****.:**.** *** **** **** .* * ***.***: :* * :** PMAGTF Y RLP AP G EP PF V K V G D K V Q K G Q V C I I E AM KL M NE I E AD Q S G T I PMAGTF Y RLP AP G EP PF V K V G D K V Q K G Q V C I I E AM KL M NE I E AD Q S G T I PMAGTF Y RLP AP G EP PF V K V G D K V Q K G Q V C I I E AM KL M NE I E AD Q S G T I PMAGTF Y RLP AP G EP PF V K V G D K V Q K G Q V C I I E AM KL M NE I E AD Q S G T I PMAGTF Y RLP AP G EP PF V K V G D K V Q K G Q V C I I E AM KL M NE I E AD Q S G T I PMAGTF Y RLP AP G EP PF V K V G D K V Q K G Q V C I I E AM KL M NE I E AD Q S G T I PMAGTF Y RLP AP G EP PF V K V G D K V Q K G Q V C I I E AM KL M NE I E AD Q S G T I PMAGTF Y RLP AP G EP PF V K V G D K V Q K G Q V C I I E AM KL M NE I E AD Q S G T I ******* ***************************	200
AM-BCCP Vernicia fordii Jatropha curcas Ricinus communis AM-BCCP Vernicia fordii Jatropha curcas Ricinus communis	AQP SYQHALLP SPPPAAP AF APP SSTSPP-APALP SPAKTNTSRTP-LKC AQP SYQHAMLP SPPLAAP AS APP SSTPPPPAPALP SPAKSSTSSHPQLKC MPP SYQHTMLP SPPPAAP AS APP SS — PPLALALPPPAKTSSSSHPPLKC MPP SYQHVMLPPPP SAAP — PPSS — AP-APALP SPAKAVTSSHPAFKC *****.:**.** *** *** *** *** *** **** *	200
AM-BCCP Vernicia fordii Jatropha curcas Ricinus communis AM-BCCP Vernicia fordii Jatropha curcas Ricinus communis AM-BCCP	AQP SYQHALLP SPPPAAP AF APP SSTSPP-APALP SPAKTNTSRTP-LKC AQP SYQHAMLP SPPLAAP AS APP SSTPPPPAPALP SPAKSSTSSHPQLKC MPP SYQHTMLP SPPPAAP AS APP SS — PPLALALPPPAKTSSSSHPPLKC MPP SYQHVMLPPPP SAAP — PPSS — AP-APALP SPAKAVTSSHPAFKC *****.:**.** *** **** **** *** *** *** *	200
AM-BCCP Vernicia fordii Jatropha curcas Ricinus communis AM-BCCP Vernicia fordii Jatropha curcas Ricinus communis AM-BCCP Vernicia fordii	AQP SYQHALLP SPPPAAP AF AP AF AP PS ST SPP-AP ALP SPAKTNT SRTP-LKC AQP SYQHAMLP SPPLAAP AS APP SS TPPP AP ALP SPAK ST SSHPQLKC MPP SYQHTMLP SPPPAAP AS APP SS — PP LALALPPPAKTSSS SHPPLKC MPP SYQHVMLPPPP SAAP — PP SS — AP - AP ALP SPAK AVT SSHPAF KC *****.:**.** *** **** **** .****.***::*** ::*** PMAGTF YRLP AP GEPPF VKVGD KVQKGQ VVCIIE AMKLMNE IE AD QSGTI PMAGTF YR CP AP GEPPF VKVGD KVQKGQ VVCIIE AMKLMNE IE AD QSGTI PMAGTF YR NP AP GEPPF VKVGD KVQKGQ VVCIIE AMKLMNE IE AD QSGTI PMAGTF YR NP AP GEPPF VKVGD KVQKGQ VVCIIE AMKLMNE IE AD QSGTI PMAGTF YR NP AP GEPPF VKVGD KVQKGQ VVCIIE AMKLMNE IE AD QSGTI ******* *****************************	200
AM-BCCP Vernicia fordii Jatropha curcas Ricinus communis AM-BCCP Vernicia fordii Jatropha curcas Ricinus communis AM-BCCP Vernicia fordii Jatropha curcas	AQP SYQHALLP SPPPAAP AFAPP SSTSPP-APALP SPAKTNTSRTP-LKC AQP SYQHAMLP SPPLAAP ASAPP SSTPPPPAPALP SPAKSSTSSHPQLKC MPP SYQHTMLP SPPPAAP ASAPP SS — PPLALALPPPAKTSSSSHPPLKC MPP SYQHVMLPPPP SAAP — PPSS — AP-APALP SPAKAVTSSHPAFKC *****.:**.** *** **** *** *** *** *** **	200
AM-BCCP Vernicia fordii Jatropha curcas Ricinus communis AM-BCCP Vernicia fordii Jatropha curcas Ricinus communis AM-BCCP Vernicia fordii Jatropha curcas Ricinus communis	AQP SYQHALLP SPPPAAP AF APP SSTSPP-APALP SPAKTNTSRTP-LKC AQP SYQHAMLP SPPLAAP AS APP SSTPPPPAPALP SPAKSSTSSHPQLKC MPP SYQHTMLP SPPPAAPASAPPSS—PPLALALPPPAKTSSSSHPPLKC MPP SYQHVMLPPPP SAAP—PPSS—AP-APALP SPAKAVTSSHPAFKC *****.:**.** *** **** **** ****.***::*** PMAGTFYRLP APGEPPFVKVGDKVQKGQVVCIIE AMKLMNE IE ADQ SGTI PMAGTFYRCP APGEPPFVKVGDKVQKGQVVCIIE AMKLMNE IE ADQ SGTI PMAGTFYRNP APGEPPFVKVGDKVQKGQVVCIIE AMKLMNE IE ADQ SGTI PMAGTFYRNP APGEPPFVKVGDKVQKGQVVCIIE AMKLMNE IE ADQ SGTI PMAGTFYRNP APGEPPFVKVGDKVQKGQVVCIIE AMKLMNE IE ADQ SGTI ******* *****************************	200

Figure 2. Amino acid homology comparison of the BCCP subunit in *Aleurites moluccana* and other higher plant species. *Fully conserved residue.

Genetics and Molecular Research 14 (3): 9922-9931 (2015)

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9927

W.Y. Xuan et al.



Figure 3. Hydropathicity of the BCCP subunit in Aleurites moluccana.

Expression analysis of AM-accB and changes in oil content during seed development

The development period of *A. moluccana* seeds can be as long as 6 months in southern China. During the initial development stage, no endosperm is formed in the seeds of *A. moluccana* in March and April. Thus, we determined the lipid content of seeds from May onwards. The results show that the lipid content of seeds gradually increases during the process of development and maturation. Both the oil content and its accumulation rate were low in the early stage of development, and the accumulation rate of oil reached its highest level during the middle stage of development (June to August), with the oil content in dry kernels reaching 67.54% during the maturation stage (Figure 4). The expression pattern of the *AM-accB* gene during *A. moluccana* endosperm development was very similar to that of the oil accumulation rate. The highest expression level of *AM-accB* was observed during the middle stage of development (July) and expression during the maturation stage was higher than that during the early stage of development (Figure 5).



Month

Figure 4. Variation in the lipid content of kernels at different stages during the development of *Aleurites moluccana* seeds.

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9928



Figure 5. Expression of the AM-accB gene at different stages during the development of the endosperm in *Aleurites* moluccana.

DISCUSSION

The BCCP subunit is one of four subunits of ACCase. In many plants, the cDNA sequence length of the BCCP subunit gene ranges from 1013 bp to 1480 bp, the deduced amino acid sequences are 258 to 286 amino acids in length, and the amino acid sequences have high identity between plants, ranging from 70 to 86% (Gu et al., 2011; Cui et al., 2012; Gong et al., 2013). Previous research has shown that the expression patterns of the BCCP subunit gene are very similar to that of the changes in oil accumulation during seed development (Cui et al., 2012), but to date there has been no report on the structural characteristics of the BCCP subunit gene and its expression patterns in *A. moluccana*. Therefore, the results of this study will be useful for further exploring the sequence polymorphisms of the BCCP subunit gene and developing molecular markers related to seed oil content in *A. moluccana*.

Although there are a number of methods for cloning the full-length coding sequence of plant genes, such as shotgun, *in silico* cloning, and chromosome walking, the homology cloning strategy combined with the RACE technique adopted in our study is one of the most simple, effective, and low cost.

The expression profile of the BCCP subunit gene in *A. moluccana* during the process of seed development and maturation is similar to that of its homologues in other woody plants, such as *J. curcas*, *V. fordii*, and *C. oleifera* (Gu et al., 2011; Cui et al., 2012; Zhou , 2013). Fatty acids are first synthesized and then changed into oil during the lipid synthesis process of plant seed development. During plant seed development, the expression peak of the ACCase subunit genes (accA, accB, accC, accD) appears in the fastest period of oil accumulation. In addition to accC, the transcriptional level of the other three genes was approximately 6 to 12-fold higher than that in the early stage of seed development (Chen et al., 2009; Rogalski and Carrer, 2011). The highest expression of genes involved in fatty acid synthesis appeared approximately one month earlier than the fastest period of oil content increase in *C. oleifera* (Zhou, 2013). These results suggest that the BCCP subunit gene may play an important role in the fatty acid synthesis process. Further research into the relationship between gene expression and single nucleotide polymorphisms (SNPs) in the BCCP subunit gene and the content change of fatty acid during *A. moluccana* seed development is required.

Genetics and Molecular Research 14 (3): 9922-9931 (2015)

Conflicts of interest

The authors declare no conflict of interest.

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Genetics and Molecular Research 14 (3): 9922-9931 (2015)