

# Genetic structure in fragmented populations of *Solanum lycocarpum* A. St.-Hil. with distinct anthropogenic histories in a Cerrado region of Brazil

T.M. Moura<sup>1</sup>, K. Martins<sup>2</sup>, P.S. Sujii<sup>3</sup>, A.M. Sebbenn<sup>4</sup> and L.J. Chaves<sup>5</sup>

<sup>1</sup>Programa de Pós-Graduação em Biologia Vegetal, Instituto de Biologia, Universidade Estadual de Campinas, Campinas, SP, Brasil
<sup>2</sup>Departamento de Biologia, Universidade Federal de São Carlos, Campus Sorocaba, Sorocaba, SP, Brasil
<sup>3</sup>Programa de Pós-Graduação em Genética e Biologia Molecular, Instituto de Biologia, Universidade Estadual de Campinas, Campinas, SP, Brasil
<sup>4</sup>Instituto Florestal de São Paulo, São Paulo, SP, Brasil
<sup>5</sup>Setor de Melhoramento Vegetal, Universidade Federal de Goiás, Goiânia, GO, Brasil

Corresponding author: T.M. de Moura E-mail: tmariamoura@gmail.com

Genet. Mol. Res. 11 (3): 2674-2682 (2012) Received November 10, 2011 Accepted April 29, 2012 Published July 10, 2012 DOI http://dx.doi.org/10.4238/2012.July.10.16

**ABSTRACT.** Solanum lycocarpum is a woody tree widely distributed in the Cerrado that reaches high population densities in disturbed environments. We examined the genetic diversity and population differentiation of six *S. lycocarpum* populations with different degrees of human disturbance in order to determine if they are negatively affected by anthropogenic activity. Three populations located in southern and three located in southeastern regions of Goiás State, Central Brazil, were genotyped with five microsatellite markers. The population located in a protected area had higher number of alleles (26) than the remaining populations (19 to 21

Genetics and Molecular Research 11 (3): 2674-2682 (2012)

alleles). It indicates that extensive and continuous areas of preserved native vegetation contribute positively to the conservation of genetic diversity, even with *S. lycocarpum* that easily adapts to disturbed environments. The three southeastern populations, although fragmented, had preserved native vegetation and were not significantly different from each other ( $\theta p = 0.002$ ). All other population pairs compared were significantly divergent ( $\theta p$  varied from 0.03 to 0.11 between pairs, P < 0.05). We found three distinct sets of allele frequencies. The three southeastern populations shared similar gene pools, as well as the two disturbed southern populations, which are secondary vegetation. The southern population located in protected area had the most dissimilar gene pool. In conclusion, populations showing a higher degree of human disturbance tends to show a larger population differentiation than expected from the isolation by distance model, which in the current scenario of the Cerrado destruction points out to a threat to the long-term conservation of the species.

**Key words:** Wolf fruit; Genetic conservation; Population differentiation; Genetic diversity; Microsatellite marker; Gene flow

# INTRODUCTION

The Cerrado is the second largest Brazilian biome, and it is biologically the richest savannah in the world. Around 57% of the original Cerrado vegetation has been completely destroyed, and half of the remaining areas have been extensively degraded. The annual rate of deforestation is estimated to be 1.5%, which corresponds to three million hectares annually, a rate ten times higher than that estimated for the Atlantic Forest (Machado et al., 2004). Because of endemism, where 44% of vascular plant species are endemic, and due to the current rate of devastation, the Cerrado is considered one of the 25 biodiversity hotspots for global conservation (Myers et al., 2000). Up to 3% of the Cerrado range is in protected areas (Aguiar et al., 2004), and the distribution of these areas is not homogeneous throughout the biome. As a consequence, an important part of the diversity is not embedded in the national network of protected areas (Silva and Bates, 2002). Thus, one of aims for this biome's conservation is to identify priority areas for conservation and to plan the establishment of new protected areas.

The conservation of genetic diversity is a necessary precondition for the maintenance of all levels of biodiversity and is an essential component of the sustainability of populations (Boyle, 2000; Namkoong et al., 2002). The reduction in natural habitats and the subsequent spatial isolation of populations cause changes in basic evolutionary processes of the populations, such as genetic drift, selection and migration, which result in changes in genetic diversity and structure. Therefore, to maintain levels of genetic diversity and to avoid extinction, it is necessary to conserve these evolutionary processes (Namkoong et al., 2002).

In the last decade, several studies have evaluated the genetic structure in Cerrado plant species (Zucchi et al., 2003; Soares et al., 2008; Moreno et al., 2009; Collevatti et al., 2001, 2010; Telles et al., 2003, 2010; Tarazi et al., 2010; Martins et al., 2006, 2011; Moura et al., 2009, 2011; Moraes and Sebbenn, 2011). Most of them addressed the genetic diversity and population differentiation, aiming to determine the conservation status of natural populations, which would then make it possible to infer historical events related to anthropogenic

Genetics and Molecular Research 11 (3): 2674-2682 (2012)

#### T.M. Moura et al.

disturbance, gene flow between populations and the effect of ecosystem fragmentation. In small populations, genetic drift is a significant determinant of the genetic structure; unlike in large populations, where the main factor influencing the genetic structure of plant species is reproductive biology, especially because reproductive biology determines the mating systems and the extent of the gene flow among populations.

Solanum lycocarpum A. St.-Hil is an interesting plant species for the study of population genetic structure geared towards revegetation: 1) the species is widely distributed in the Cerrado; 2) the species occupies rapidly degraded environments, where it reaches high population densities; 3) the amount of knowledge regarding the reproductive biology of the species is considerable (Oliveira-Filho and Oliveira, 1988; Lombardi and Motta Jr., 1993; Courtenay, 1994; Moura et al., 2010); and 4) pollinators and seed dispersers contribute to longdistance gene flow (Martins et al., 2011). Because of these attributes, we expect no significant population differentiation. If the opposite were observed, we could infer it is a negative effect of human disturbance, as a consequence of genetic drift and/or restriction in gene flow. The establishment of secondary vegetation and the fragmentation are common types of human disturbance found in Cerrado biome, which could cause strong genetic drift and limited gene flow. We aimed to assess the genetic diversity and divergence among *S. lycocarpum* populations with different anthropogenic disturbance histories, to determine if there have been any negative effects and to contribute to the establishment of conservation plans for the Cerrado biome.

# **MATERIAL AND METHODS**

### **Study species**

*S. lycocarpum* is a woody species which produces functional male and hermaphrodite flowers in large quantities throughout the year. Large bees pollinate them (Oliveira-Filho and Oliveira, 1988). Various mammals disperse seeds over long distances, thus allowing the colonization of new environments. Since the fruits only finish their ripening process after falling to the ground (Lombardi and Motta Jr., 1993) many seeds germinate under the seed tree's canopy or are scattered over short distances by leaf-cutter ants (Courtenay, 1994). The species occurs at a low density on primary vegetation but it is able to colonize anthropogenic environments, where it reaches higher densities. Although the higher population density would imply an apparent increase in effective population size, the simple increase in the number of individuals is not synonymous with genetic sustainability, as detected by Moura et al. (2011).

# Study site and sampling

Six *S. lycocarpum* populations were studied in Goiás State, in Central Brazil. Three located in the southeast (called A, B and C) and the other three in the south (D, E and F) (Table 1). The F population has preserved vegetation and is situated in a protected area: the Parque Estadual da Serra de Caldas Novas. The remaining populations are located in fragments belonging to private ranches. Southeast populations were located in the Cerrado region encompassing areas of preserved vegetation and with low anthropogenic influence. The D and E populations consist of pastureland; however, D still retains native vegetation, and E has a predominance of signal grass (*Urochloa* spp) with few native species.

Genetics and Molecular Research 11 (3): 2674-2682 (2012)

Table 1. Description of the Solanum lycocarpum A. StHil. populations studied in Goiás State, Brazil.									
Region	Population	Municipality	Location	Vegetation condition					
Southeast	А	Bela Vista de Goiás	17°00'S; 48°46'W	Preserved Cerrado					
	В	São Miguel do Passa Quatro	17°00'S; 48°35'W	Preserved Cerrado					
	С	Orizona	17°03'S; 48°22'W	Preserved Cerrado					
South	D	Morrinhos	17°54'S; 49°11'W	Pastureland, with native vegetation					
	Е	Morrinhos	17°55'S; 49°00'W	Pastureland, with fewer native vegetation					
	F	Caldas Novas	17°46'S; 48°40'W	Preserved Cerrado					

Sixty adult individuals were sampled for each population. Genomic DNA extraction was according to Doyle and Doyle (1987), with modifications. Five polymorphic microsatellite (SSR) primers developed for *Capsicum* spp and already amplified in *S. lycocarpum* (Martins et al., 2006) were used. The conditions of polymerase chain reaction (PCR) and the characterization of loci are described by Martins et al. (2006). After amplification, the DNA fragments were separated using a 4% polyacrylamide gel and visualized after staining with silver nitrate. The details of the procedures can be found in Martins et al. (2006) and Moura et al. (2009).

## Data analysis

We estimated the total number of alleles, the number of rare alleles (frequency of occurrence less than 0.05), and the fixation index (*f*) for each population. The 95% confidence interval of the fixation index was calculated using 1000 bootstraps over loci. Estimates of other parameters of genetic diversity for these populations were previously published (Martins et al., 2006; Moura et al., 2009). In this study, the average gene diversity ( $H_E$ ) and allelic richness ( $R_s$ ) (Petit et al., 1998) were both compared in the southeastern (populations A, B and C) and southern (populations D, E and F) regions by the *G*-test (Goudet et al., 1996) with 1000 randomizations and Bonferroni's correction. All analyses were carried out using the FSTAT software (Goudet, 1995).

The genetic structure was characterized by the analysis of variance of gene frequencies (Weir, 1996), using the FSTAT software. The estimated parameters were: mean fixation index within populations (f), fixation index of the total set of populations (F) and genetic differentiation among populations ( $\theta$ p). The 95% confidence intervals were estimated by 1000 bootstraps over loci. Subsequently, we estimated the genetic differentiation between pairwise populations ( $\theta$ p). The statistical significance of genetic differentiation between populations was assessed using the *G*-test (Goudet et al., 1996) with 1000 randomizations and Bonferroni's correction. The indirect gene flow ( $N_m$ ) between pairs of populations was estimated according to Wright (1951). For these calculations,  $F_{st}$  was replaced by  $\theta$ p, as suggested by Cockerham and Weir (1993) for an unbiased estimation of gene flow.

We applied the procedure described by Slatkin (1993) to assess whether gene flow results in a pattern of isolation by distance. The approach is based on the linear relationship between gene flow  $(N_m)$  and logarithms of geographical distance between populations. As discussed by Strand et al. (1996), in the context of isolation by distance and in a stepping-stone model of gene flow, a strong negative correlation indicates that populations are in equilibrium and that the current gene flow results in a pattern of isolation by distance, where genetic differentiation between populations increases with geographic distance. The lack of correlation

Genetics and Molecular Research 11 (3): 2674-2682 (2012)

suggests that the prevailing gene flow mainly results in historical association among populations than the current inter-population one. Estimates of  $\theta$ p for each pair of populations were used to calculate the gene flow between them. The significance of the correlation between the logarithm of geographic distance and gene flow was evaluated by Mantel Z statistics (Mantel, 1967) using 1000 permutations, with the NTSYS-pc software (Rohlf, 1989). The genetic divergence estimates  $G_{\rm ST}$  (Hedrick, 2005) for the whole population were obtained through the GDA\_NT software (Degen, 2006).

The individual groups' separation was determined by a Bayesian approach with the Structure 2.3.1 software (Pritchard et al., 2000). The model used allowed for genome mixing, with the number of groups (*K*) ranging from 1 to 7, with 500,000 resampling by the MCMC (Markov Chain Monte Carlo) method, 100,000 burn-ins and 10 independent repetitions. The most likely number of groups was found using the statistics described by Evanno et al. (2005), based on the rate of change in data probability between successive *K* values. The peak in the  $\Delta K$  value can identify the *K* value that best represents the populations' structuring.

# RESULTS

The F population, located in a protected area, showed the highest number of alleles. The remaining populations had similar numbers of alleles, between 19 and 21 (Table 2). All populations had rare alleles, and these accounted for 33 and 43% of the total number of alleles for the southeastern and southern populations, respectively. We found that allelic richness ( $R_s$ ) and gene diversity ( $H_E$ ) were not significantly different between regions (Table 2). Inbreeding was observed only in the A population, judging by the confidence interval (Table 2).

	Southeast		Southern			
А	В	С	D	Е	F	
58	60	60	56	60	60	
19	20	21	19	18	26	
06	06	08	08	08	11	
0.214	-0.006	0.077	0.017	-0.090	-0.176	
(0.062 to 0.653) 0.348 <sup>NS</sup>	(-0.065 to 0.044)	(-0.092 to 0.1688)	(-0.157 to 0.122) 0.341 <sup>NS</sup>	(-0.275 to 0.051)	(-0.500 to 0.097)	
((	A 58 19 06 0.214 0.062 to 0.653) 0.348% 2 0.1%	A         B           58         60           19         20           06         06           0.214         -0.006           0.062 to 0.653)         (-0.065 to 0.044)           0.348 <sup>NS</sup> -2.0 <sup>NS</sup>	A         B         C           58         60         60           19         20         21           06         06         08           0.214         -0.006         0.077           0.062 to 0.653)         (-0.065 to 0.044)         (-0.092 to 0.1688)           0.348 <sup>NS</sup> -2.01 <sup>NS</sup> -2.01 <sup>NS</sup>	A         B         C         D           58         60         60         56           19         20         21         19           06         06         08         08           0.214         -0.006         0.077         0.017           0.062 to 0.653)         (-0.055 to 0.044)         (-0.092 to 0.1688)         (-0.157 to 0.122) $0.348^{NS}$ $0.341^{NS}$ $416^{NS}$	A         B         C         D         E           58         60         60         56         60           19         20         21         19         18           06         06         08         08         08           0.214         -0.006         0.077         0.017         -0.090           0.062 to 0.653)         (-0.055 to 0.044)         (-0.092 to 0.1688)         (-0.157 to 0.122)         (-0.275 to 0.051)           0.348 <sup>NS</sup> 41 (NS)         41 (NS)         41 (NS)         41 (NS)	

 $\hat{f}$  = fixation index with 95% confidence interval (95%CI) obtained by 1000 bootstraps over loci;  $\hat{H}_{\rm E}$  = gene diversity;  $R_{\rm S}$  = allelic richness. NS = not significant, P > 0.05 by the *G*-test (Goudet et al., 1996).

By determining the genetic divergence between pairs of populations (Table 3), we observed that the southeastern C population was more similar to the southern F population than to its geographically closer neighbors (D and E). This shows that F was the most genetically diverse among the populations from the same region, although the genetic divergence between populations D and E ( $\theta p = 0.033$ ) was also significantly different from zero (P < 0.05). With the exception of the comparisons between populations A, B and C, all other population pairs compared were significantly divergent (Table 3).

each pair of populations.									
Population	А	В	С	D	Е	F			
A	-	-0.003 <sup>NS</sup>	0.005 <sup>NS</sup>	0.114*	0.076*	0.098*			
В	19.5	-	0.004 <sup>NS</sup>	0.115*	0.076*	0.096*			
С	43.0	23.7	-	0.095*	0.052*	0.058*			
D	109.0	118.3	128.0	-	0.033*	0.105*			
E	104.5	110.7	117.2	19.5	-	0.080*			
F	85.5	85.3	85.5	56.7	39.0	-			

**Table 3.** Genetic divergence  $(\hat{\theta}_p)$  (above the diagonal) and geographic distance in km (below the diagonal) for each pair of populations.

NS = not significant and \* = P < 0.05 by the *G*-test (Goudet et al., 1996).

Three distinct sets of allele frequencies were observed (K = 3) (Figure 1). Populations A, B, and C shared similar gene pools, with the same being observed for the D and E populations. The F population was the one with the most dissimilar gene pool.



Figure 1. Genetic structure in six populations (A-F) of Solanum lycocarpum A. St.-Hil.

The isolation by distance test showed a moderate negative correlation yet statistically significant between gene flow and logarithms of geographic distances between pairs of populations. Only 55% of gene flow variation was explained by the variation in geographical distance. This was mainly due to the high divergence between the F population and other populations in the southern region (Table 3). A strong correlation would indicate that populations are at equilibrium between genetic drift and gene flow and that the actual gene flow results in a pattern of isolation by distance, where genetic differentiation between populations increases with geographic distance. The correlation was moderate and significant (r = -0.745; P = 0.011) showing a trend toward isolation by the distance model and that there was no balance between genetic drift and gene flow.

## DISCUSSION

The highest number of alleles was observed in the population located in Parque Estadual de Serra de Caldas Novas (F), a protected area. This is an indication that extensive and continuous areas of preserved native vegetation contributes positively to the conservation of genetic diversity, even with *S. lycocarpum*, which easily adapts to degraded environments as noted by Moura et al. (2011). In this study, we found that populations whose native vegetation is preserved (populations A, B and C), but fragmented, had fewer alleles than the population located in a protected area, containing continuous vegetation.

The fixation index of the total set of populations (F = 0.07) and average fixation index within populations (f = 0.002) did not differ from zero; which indicates that a major portion of

Genetics and Molecular Research 11 (3): 2674-2682 (2012)

#### T.M. Moura et al.

the total inbreeding is due to population subdivision, and thus, it is a result of genetic drift, and restricted gene flow between populations. Population differentiation was low ( $\theta p = 0.068$ ) but significant. The population differentiation among southeastern populations (A, B and C) ( $\theta p$ = 0.002) was not statistically different from zero. This indicates that they consisted of a single panmictic population prior to the Cerrado's fragmentation. Since nowadays native vegetation is discontinuous in the region, they can be considered subpopulations. In contrast, the divergence among the southern populations (D, E and F) was significant and greater than population differentiation in the total set of populations ( $\theta p = 0.076$ ), showing that these populations are heterogeneous among themselves. Contrary to our expectations the population differentiation estimated according to Hedrick (2005) ( $G_{ST} = 0.041$ ) was lower than the one estimated by the Weir and Cockerham (1984) statistics. This was probably due to the low polymorphism of our microsatellite loci, in contrast to what is generally observed for this type of marker. The greatest genetic divergence between the southern and southeastern (A, B and C) populations might have been due to their particular anthropogenic history. It is believed that, historically, they had suffered less anthropogenic influence than the D and E populations. This assumption was corroborated by the vegetation's appearance; D and E populations are degraded areas, possibly, being secondary type vegetation.

We noted that each pool of alleles was related to an environmental condition. The F population is situated in a protected area and had a particular set of genes, and the D and E populations apparently have similar anthropogenic histories and share a similar gene pool, the same being observed for the A, B and C populations. Our results show that even species that easily occupy degraded environments, such as *S. lycocarpum*, have experienced negative effects due to vegetation fragmentation. The significant genetic divergence among populations, as occurred in the southern part of the state where the environment is more anthropogenic, shows that the populations are losing connectivity, probably due to reduced number of populations and restriction in foraging by their pollinators and seed dispersers.

Although studies with the Cerrado's woody species have recently been published, there are still gaps in our knowledge of the genetic diversity of the Cerrado's species, which must be filled. Few studies, for example, have assessed mating systems in detail and contemporary pollen flow (see Collevatti et al., 2010; Moraes and Sebbenn, 2011). Although virtually all authors argue about the risks of the rapid rate of deforestation, the effect of habitat fragmentation has not been systematically evaluated. There is a predominant use of indirect approaches for estimating historical gene flow, one reason being that biome fragmentation is recent. Only two studies (Collevatti et al., 2003; Martins et al., 2011) used a comparative approach with nuclear and chloroplast markers to assess the relative contribution of pollen and seeds to total gene flow. Only studies by Moura et al. (2009, 2011) compared populations in fragmented and continuous vegetation. It is also worth emphasizing the remarkable territorial extent of the biome and the need for studies covering larger geographic ranges. The majority of studies were conducted in Goiás State and the Distrito Federal, while the States of Mato Grosso, Mato Grosso do Sul, Tocantins, and São Paulo have received little attention.

# ACKNOWLEDGMENTS

We are thankful to the staff of LARGEA (Laboratório de Reprodução e Genética de Espécies Arbóreas, ESALQ/USP), and to "Espaço da escrita" translator service at UNICAMP.

Genetics and Molecular Research 11 (3): 2674-2682 (2012)

Research supported by Fundação de Amparo à Pesquisa do Estado de São Paulo (FAPESP/ Brazil). K. Martins received a PhD fellowship from Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq/Brazil).

## REFERENCES

- Aguiar LMS, Machado RB and Marinho-Filho JA (2004). Diversidade Biológica do Cerrado. In: Cerrado: Ecologia e Caracterização (Aguiar LMS and Camargo AJA, eds.). EMBRAPA Cerrados, Planaltina, 17-40.
- Boyle TJ (2000). Criteria and Indicators for the Conservation of Genetic Diversity. In: Forest Conservation Genetics: Principles and Practice (Young A, Boshier D and Boyle T, eds.). CSIRO Publishing, Collingwood, 239-251.
- Cockerham CC and Weir BS (1993). Estimation of gene flow from F-statistics. Evolution 47: 855-863.
- Collevatti RG, Grattapaglia D and Hay JD (2001). Population genetic structure of the endangered tropical tree species *Caryocar brasiliense*, based on variability at microsatellite loci. *Mol. Ecol.* 10: 349-356.
- Collevatti RG, Grattapaglia D and Hay JD (2003). Evidences for multiple maternal lineages of *Caryocar brasiliense* populations in the Brazilian Cerrado based on the analysis of chloroplast DNA sequences and microsatellite haplotype variation. *Mol. Ecol.* 12: 105-115.
- Collevatti RG, Estolano R, Garcia SF and Hay JD (2010). Short-distance pollen dispersal and high self-pollination in a bat-pollinated neotropical tree. *Tree Genet. Genomes* 6: 555-564.
- Courtenay O (1994). Conservation of the maned wolf: fruitful relationships in a changing environment. *Canid News* 2: 41-43.
- Degen B (2006). GDA\_NT 2006. Genetic Data Analysis and Numerical Tests. Institute for Forest Genetics, Grosshansdorf. Doyle JJ and Doyle JL (1987). A rapid DNA isolation procedure for small quantities of fresh leaf tissue. *Phytochem. Bull.* 19: 11-15.
- Evanno G, Regnaut S and Goudet J (2005). Detecting the number of clusters of individuals using the software STRUCTURE: a simulation study. *Mol. Ecol.* 14: 2611-2620.
- Goudet J (1995). FSTAT (version 1.2): a computer program to calculate F-statistics. J. Hered. 86: 485-486.
- Goudet J, Raymond M, de MT and Rousset F (1996). Testing differentiation in diploid populations. *Genetics* 144: 1933-1940.
- Hedrick PW (2005). A standardized genetic differentiation measure. Evolution 59: 1633-1638.
- Lombardi JA and Motta JC Jr (1993). Seed dispersal of *Solanum lycocarpum* St.Hil. (Solanaceae) by the maned wolf, *Chrysocyon brachyurus* Illiger (Mammalia, Canidae). *Cienc. Cult.* 45: 126-127.
- Machado RB, Ramos Neto MB, Pereira PGP, Caldas EF, et al (2004). Estimativas de Perda da Área do Cerrado Brasileiro. Relatório Técnico. Conservation International, Brasília.
- Mantel N (1967). The detection of disease clustering and a generalized regression approach. Cancer Res. 27: 209-220.
- Martins K, Chaves LJ, Buso GSC and Kageyama PY (2006). Mating system and fine-scale spatial genetic structure of Solanum lycocarpum St.Hil. (Solanaceae) in Brazilian Cerrado. Conserv. Genet. 7: 957-969.
- Martins K, Chaves LJ, Vencovsky R and Kageyama PY (2011). Genetic structure based on nuclear and chloroplast microsatellite loci of Solanum lycocarpum A. St.-Hil. (Solanaceae) in Central Brazil. Genet. Mol. Res. 10: 665-677.
- Moraes MLT and Sebbenn AM (2011). Pollen dispersal between isolated trees in the Brazilian Savannah: a case study of the neotropical tree *Hymenaea stigonocarpa*. *Biotropica* 43: 192-199.
- Moreno MA, Tarazi R, Ferraz EM, Gandara FB, et al. (2009). Estrutura genética espacial em populações de *Hymenaea* stigonacarpa Mart. ex Hayne mediante a utilização de marcadores microssatélites cloroplastitidais. Sci. For. 37: 513-523.
- Moura TM, Sebbenn AM, Chaves LJ, Coelho ASG, et al. (2009). Diversidade e estrutura genética espacial em populações fragmentadas de *Solanum spp.* do cerrado, estimadas por meio de locos microssatélites. *Sci. For.* 37: 143-150.
- Moura TM, Oliveira GCX and Chaves LJ (2010). Correlation between flowering, fructification and environmental variables in *Solanum lycocarpum* A. St.-Hil, Solanaceae. *Biosci. J.* 26: 457-462.
- Moura TM, Sebbenn AM, Martins K, Moreno MA, et al. (2011). Allelic diversity in populations of *Solanum lycocarpum* A. St.-Hil (Solanaceae) in a protected area and a disturbed environment. *Acta Bot. Bras.* 25: 937-940.
- Myers N, Mittermeier RA, Mittermeier CG, Fonseca GAB, et al. (2000). Biodiversity hotspots for conservation priorities. *Nature* 403: 853-858.
- Namkoong G, Boyle TJB, El-Kassaby YA, Palmberg-Lerche C, et al (2002). Criteria and Indicators for Sustainability Forest Management: Assessment and Monitoring of Genetic Variation. Food and Agricutural Organization, Rome.
- Oliveira-Filho AT and Oliveira LCA (1988). Biologia floral de uma população de *Solanum lycocarpum* St.-Hil. (Solanaceae) em Lavras, MG. *Rev. Bras. Bot.* 11: 23-32.

Genetics and Molecular Research 11 (3): 2674-2682 (2012)

#### T.M. Moura et al.

- Petit RJ, El Mousadik A and Pons O (1998). Identifying populations for conservation on the basis of genetic markers. *Conserv. Biol.* 12: 844-855.
- Pritchard JK, Stephens M and Donnelly P (2000). Inference of population structure using multilocus genotype data. *Genetics* 155: 945-959.
- Rohlf FJ (1989). NTSYS-pc: Numerical Taxonomy and Multivariate Analysis System. Exeter publisher, New York.
- Silva JMC and Bates JM (2002). Biogeographic patterns and conservation in the South American Cerrado: a tropical savanna hotspot. *BioScience* 52: 225-234.

Slatkin M (1993). Isolation by distance in equilibrium and non-equilibrium populations. Evolution 47: 264-279.

- Soares TN, Chaves LJ, de Campos Telles MP, Diniz-Filho JA, et al. (2008). Landscape conservation genetics of *Dipteryx* alata ("baru" tree: Fabaceae) from Cerrado region of central Brazil. *Genetica* 132: 9-19.
- Strand AE, Milligan BG and Pruitt CM (1996). Are populations islands? Analysis of chloroplast DNA variation in *Aquilegia. Evolution* 50: 1822-1829.
- Tarazi R, Moreno MA, Gandara FB, Ferraz EM, et al. (2010). High levels of genetic differentiation and selfing in the Brazilian cerrado fruit tree *Dipteryx alata* Vog. (Fabaceae). *Genet. Mol. Biol.* 33: 78-85.
- Telles MPC, Valva FD, Bandeira LF and Coelho ASG (2003). Caracterização genética de populações naturais de araticunzeiro (*Annona crassiflora* Mart. Annonaceae) no Estado de Goiás. *Rev. Bras. Bot.* 26: 123-129.
- Telles MPC, Silva SP, Ramos JR, Soares TN, et al. (2010). Estrutura genética em populações naturais de *Tibouchina* papyrus em áreas de campo rupestre no cerrado. *Rev. Bras. Bot.* 33: 291-300.
- Weir BS (1996). Genetic Data Analysis II. Sinauer Associates, Sunderland.
- Weir BS and Cockerham CC (1984). Estimating F-statistics for the analysis of population structure. *Evolution* 38: 1358-1370.
- Wright S (1951). The genetical structure of populations. Ann. Eugenics 15: 323-354.
- Zucchi MI, Brondani RPV, Pinheiro JB, Chaves LJ, et al. (2003). Genetic structure and gene flow in *Eugenia dysenterica* DC. in the Brazilian Cerrado utilizing SSR markers. *Genet. Mol. Biol.* 26: 449-457.

Genetics and Molecular Research 11 (3): 2674-2682 (2012)