

Distinct linear hydrocarbon profiles and chemical strategy of facultative parasitism among *Mischocyttarus* wasps

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ABSTRACT. Insects use pheromones as a means of chemical communication. Pheromones act on individual receptors and produce specific behavioral or physiological responses that are fundamental to intra- and interspecific recognition. The objective of this study was to evaluate the interspecific differences among the linear hydrocarbon profiles of the cuticles of 3 wasp species of the genus *Mischocyttarus*. The chemical strategy that permits an interaction among 2 of these species was also examined about their hydrocarbon profiles. The cuticular hydrocarbons present on the abdomen of each individual were extracted with hexane in an ultrasonic bath and analyzed using gas chromatography with a flame ionization detector. The results suggested that the wasp species have distinct chemical signatures as the linear

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hydrocarbons of their cuticles. However, these signatures are more alike in those with similar morphological and behavioral aspects. These similarities facilitate an interaction of facultative parasitism observed among these species, in which the invaders acquire parts of the chemical signature of the host colony, possibly as a chemical strategy to increase the likelihood that an invader will be accepted by the females of a host colony. Both invader and host have their own chemical signature changed by the first contact between the species.

Keywords: Facultative parasitism; Linear alkanes; Gas chromatography

INTRODUCTION

Social insects exhibit labor division, generation overlap, and cooperative care of offspring (Ross and Matthews, 1991). The completion of multiple tasks in their colonies requires interactions between individuals. Therefore, chemical communication mechanisms via pheromones have been developed during evolution. These compounds act on the individual receiver, producing specific behavioral or physiological responses important in both intra- and interspecific communication (Gullan and Cranston, 2007).

Pheromones have key functions in many aspects of the insect life cycle and are generally divided into 2 types: light and volatile substances secreted by glands and hydrocarbon chains in the cuticle (Howard, 1993). The cuticular hydrocarbons include a wide variety of compounds such as unsaturated hydrocarbons and a series of linear and methyl-branched saturated hydrocarbons (Sledge et al., 2001).

The variation in the chemical composition of the cuticle allows nestmate and non-nestmate recognition (Gamboa, 1996). Insects can also distinguish the larvae of their own colonies (Cotoneschi et al., 2007), workers according to their function (Abdalla et al., 2003; Antonialli Jr. et al., 2008), colonial signatures (Tannure-Nascimento et al., 2007), and even various species (Antonialli Jr. et al., 2007). Furthermore, Espelie and Hermann (1990) have noted that the elements of nest substrates contain hydrocarbons similar or complementary to those that cover the cuticle of adults insect, and they also have important roles in intra- and interspecific recognition.

Polistinae eusocial wasps live in colonies founded by a single female or by more than one female with independent foundations (Ross and Matthews, 1991). Instead of initiating a colony, however, some species can take possession of other colonies owing to the low defensive motivation of the foundresses (Nonacs and Reeve, 1995). Intra- and interspecific facultative parasitism has been described in wasps of the Polistinae family: *Vespula, Polistes,* and *Mischocyttarus* (Tinault and Ruano, 1999). Specific cases of intraspecific parasitism have been described in *Mischocyttarus flavitarsus* (Litte, 1979) and *M. mexicanus* (Clouse, 1995). In the behavior of social parasitism, beyond exhibiting similar morphological and behavioral aspects to the host, invader species also rub the posterior part of their abdomens against the comb to acquire part of the "scent" of the nest substrate, thereby imitating the chemical signature of the colony and in turn being accepted by the host individuals (Cervo and Dani, 1996; Gamboa, 1996; Lorenzi et al., 2011).

M. consimilis Zikán 1949 is a Neotropical wasp that has been studied recently (Montagna et al., 2009, 2010; Torres et al., 2011). The biological aspects of *M. cerberus* Richards

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(1940) have been described by Giannotti (1998, 1999), Silva and Noda (2000), Filho Costa et al. (2011), and Togni and Giannotti (2007, 2008, 2010). Females from these species have similar nests and morphologies, differing only in the coloration pattern of the abdomen (Giannotti, 1999; Montagna et al., 2010). The morphological aspects of *M. latior* females differ from those of *M. consimilis* and *M. cerberus* (Garcete-Barrett, 2001; Henrique-Simões et al., 2011). We aimed to evaluate the interspecific differences among the linear hydrocarbon profiles of the cuticle of these 3 species of social wasps from the *Mischocyttarus* genus. Among them, 2 display a facultative parasitic interaction in all areas evaluated in which they co-occur. We also evaluated whether these species use mimicry of the chemical signature as strategy for successful facultative parasitic interactions.

MATERIAL AND METHODS

Material collection

To evaluate the differences in cuticular hydrocarbon profiles among wasp species, we collected colonies from various localities in the Mato Grosso do Sul State region (Bodoquena, 20°32'19"S, -56°42'54"W; Baytaporã, 22°13'16"S, 54°48'20"W; Dourados, 22°13'16"S, 54°48'20"W) and one region of São Paulo State (Piracicaba, -22°42'30"S, 47°38'01"W) without any interaction between the selected colonies. Three colonies were of *M. cerberus*, 4 of *M. consimilis*, and 2 of *M. latior*. By evaluating different colonies of different populations, we expected to obtain a most reliable assessment of the profile of hydrocarbons, so we have like a medium profile of each species. The colonies were collected by placing a dark plastic package over the nest and releasing the pedicel. The wasps were anesthetized via chilling on ice, and the abdomens of 4 to 10 individuals were extracted depending on the population of each colony. To evaluate the effects of interactions between *M. cerberus* and *M. consimilis* wasps during facultative parasitism, the cuticular hydrocarbon profiles of all of the individuals of 2 colonies of *M. cerberus* invaded by *M. consimilis* were analyzed.

Chemical and statistical analysis

The cuticular hydrocarbons of 75 abdomens were extracted in 2 mL hexane (high-performance liquid chromatography grade, Vetec - Química Fina Ltda., Rio de Janeiro, RJ, Brazil) in an ultrasonic bath for 30 min. This process was performed in duplicate. In sequence, the 2 fractions were united and the solvent dried. The dried extract was dissolved in 100 μ L hexane for analysis using gas chromatography with a flame ionization detector (Focus, GC, Thermo Scientific, San Jose, CA, USA). The gas chromatograph with flame ionization detector was equipped with an OV-5 capillary column (Ohio Valley Specialty Company, Marietta, OH, USA) with a composition of 5% phenyl-dimethyl-polysiloxane in a fused silica capillary with dimensions of 30 m in length x 0.25 mm in diameter x 0.25- μ m film thickness. Nitrogen was used as the carrier gas at a constant pressure of 0.8 bar. The analyses were performed in splitless mode with 1- μ L injection, injector temperature of 250°C, and detector temperature of 320°C. The oven was programmed to reach a final temperature of 320°C, starting from an initial temperature of 50°C and rising at a rate of 5°C/min to 85°C, then rising at a rate of 8°C/min to 300°C, which was held for 15 min, and then raised to 320°C at a rate of 10°C/min and held for 35 min.

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The chromatograms were registered using the ChromQuest 5.0 program and analyzed with the Workstation Chrom Data Review program. The chemical compounds were identified based on linear hydrocarbon standards of C_7 to C_{40} (4 µg/mL in hexane) through comparison with retention times. The concentration of each compound was calculated using relative area, in percentage, referring to the peak areas of alkanes C_7 to C_{40} .

The differentiation analysis of the species was completed using the peak areas of the corresponding C_{15} to C_{30} hydrocarbons, which are common to all of the species. The differentiation was carried out using the stepwise discriminate function analysis of Statistica 7 and indicated by selecting a set of variables that best differentiate (case has difference) the groups analyzed (Quinn and Keough, 2002). In this analysis, the Wilk's lambda statistic is used as a measurement of the difference between the groups, in which values close to 0 indicate that the groups do not overlap, whereas values close to 1 indicate high overlap among groups and a lack of significant difference.

RESULTS AND DISCUSSION

Figure 1 shows the discriminate analysis of the peak areas of linear alkanes identified in the cuticular extract of worker abdomens of *M. consimilis*, *M. cerberus*, and *M. latior*. The dispersion graphic shows that the chemical compositions of the hydrocarbon profiles of the 3 species are distinct. In fact, the values of Wilk's lambda of 0.017 and F of 23.064 demonstrate significant differences between the hydrocarbon profiles (P < 0.000), with canonical root 1 explaining 70% and, along with root 2, 100% of the data. These results reinforce the importance of hydrocarbon profiles in the recognition of signals and species distinction, corroborating the findings of Howard and Blomquist (2005).



Figure 1. Dispersion diagram of discriminate analysis results showing the two canonical roots of differentiation of 3 species of wasps of the genus *Mischocyttarus* based on the profiles of cuticular hydrocarbons.

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Table 1 lists the linear alkanes present or absent in the hydrocarbon profiles of the 3 wasp species, given that alkanes C_{35} to C_{40} were absent in all of the species studied. Hydrocarbons C_{15} to C_{30} were the most important in distinguishing the species because they are present in each but in different concentrations (see Table 1). The results show that the profile of *M. latior* is more distinct than that of the other 2 species (see Figure 1), mainly owing to the presence of a large concentration of *n*-heptacosane (C_{27}). Greater overlap was found between *M. consimilis* and *M. cerberus*. These species also share similar morphological and behavioral characteristics (Giannotti, 1998, 1999; Silva and Noda, 2000; Togni and Giannotti, 2007, 2008, 2010; Montagna et al., 2009, 2010; Filho Costa et al., 2011; Torres et al., 2011). *N*-nonacosane (C_{29}) and *n*-triacontane (C_{30}), in the sequence, identified the *M. consimilis* and *M. cerberus* species because these compounds are found in high concentrations in these species (see Table 1).

Table 1. Identification of hydrocarbons present or absent in the hydrocarbon profiles and the statistical values of the discriminate analysis of the peaks corresponding to the most significant alkanes for species differentiation followed by average concentration values in percentage of area.

| łC | Nomenclature | Presence or absence | | | Wilk's lambda | Р | Canonical coefficients | | % Relative area | | |
|-----|--------------------|---------------------|-------------|-----------|------------------|-------------|---------------------------|--------|-------------------|-------------------|------------------|
| | | M. consimilis | M. cerberus | M. latior | - | | Root 1 | Root 2 | M. consimilis | M. cerberus | M. latior |
| 27 | n-heptane | 1 | 1 | 0 | - | - | - | - | - | - | - |
| 8 | n-octane | 1 | 1 | 0 | - | - | - | - | - | - | - |
| 2 | n-nonane | 1 | 1 | 1 | - | - | - | - | - | - | - |
| 210 | n-decane | 0 | 1 | 0 | - | - | - | - | - | - | - |
| 211 | n-undecane | 1 | 1 | 1 | - | - | - | - | - | - | - |
| 212 | n-dodecane | 0 | 0 | 1 | - | - | - | - | - | - | - |
| 213 | n-tridecane | 0 | 1 | 0 | - | - | - | - | - | - | - |
| 214 | n-tetradecane | 1 | 1 | 1 | - | - | - | - | - | - | - |
| 15 | n-pentadecane | 1 | 1 | 1 | 0.03 | <0.00 | 0.21 | -1.53 | 0.11 ± 0.07 | 0.44 ± 0.46 | 0.06 ± 0.02 |
| 216 | n-hexadecane | 1 | 1 | 1 | 0.02 | ns | | | | | |
| 17 | n-heptadecane | 1 | 1 | 1 | 0.02 | < 0.02 | -0.66 | 1.03 | 0.48 ± 0.24 | 0.83 ± 0.92 | 0.16 ± 0.04 |
| 18 | n-octadecane | 1 | 1 | 1 | 0.02 | ns | | | | | |
| 19 | n-nonadecane | 1 | 1 | 1 | 0.02 | ns | | | | | |
| 20 | n-eicosane | 1 | 1 | 1 | | | | | | | |
| 21 | n-heneicosane | 1 | 1 | 1 | 0.02 | < 0.05 | -0.17 | -0.58 | 6.24 ± 6.72 | 5.55 ± 11.71 | 0.06 ± 0.02 |
| 22 | n-docosane | 1 | 1 | 1 | | | | | | | |
| 23 | n-tricosane | 1 | 1 | 1 | 0.02 | ns | | | | | |
| 24 | n-tetracosane | 1 | 1 | 1 | 0.02 | ns | | | | | |
| 24 | n-pentacosane | 1 | 1 | 1 | 0.02 | ns | | | | | |
| 26 | n-hexacosane | 1 | 1 | 1 | 0.02 | < 0.01 | 1.00 | -1.40 | 0.71 ± 1.18 | 3.37 ± 3.79 | 3.48 ± 0.89 |
| 27 | n-heptacosane | 1 | 1 | 1 | 0.03 | $<\!\!0.00$ | 1.10 | 0.51 | 17.59 ± 11.70 | 12.41 ± 9.40 | 73.54 ± 11.3 |
| 28 | n-octacosane | 1 | 1 | 1 | 0.02 | ns | | | | | |
| 29 | n-nonacosane | 1 | 1 | 1 | 0.02 | < 0.02 | -0.56 | 0.28 | 46.32 ± 22.00 | 39.13 ± 18.45 | 11.67 ± 8.01 |
| 30 | n-triacontane | 1 | 1 | 1 | 0.02 | < 0.02 | 0.36 | -0.58 | 1.74 ± 1.17 | 13.42 ± 6.78 | 0.66 ± 0.27 |
| 31 | n-hentriacontane | 1 | 1 | 1 | - | - | - | - | - | - | - |
| 32 | n-dotriacontane | 1 | 1 | 1 | - | - | - | - | - | - | - |
| 33 | n-tritriacontane | 1 | 1 | 0 | - | - | - | - | - | - | - |
| 33 | n-tetratriacontane | e 1 | 1 | 0 | - | - | - | - | - | - | - |

HC = hydrocarbons; ns = non-significant.

Dapporto et al. (2006) have also found high concentrations of linear alkanes with the carbon chains of C_{23} to C_{33} in *Ropalidia opifex*. In *Polistes satan*, the linear alkanes of *n*-heptacosane (C_{27}), *n*-octacosane (C_{28}), and *n*-tritriacontane (C_{33}) are more significant than methyl-branched alkanes in the separation of colonies (Tannure-Nascimento et al., 2007).

The results demonstrate that *M. consimilis* and *M. cerberus* have distinct hydrocarbon profiles (Figure 2), that a clear separation of the groups is present, and that the significant

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differences among the linear hydrocarbon profiles of 2 of the species were detected in the canonical discrimination analysis (Wilk's lambda = 0.036; F = 8.792; P < 0.000). This separation was determined by the linear alkanes C_{15} , C_{19} , C_{23} , C_{26} , C_{27} , C_{29} , and C_{30} (Table 2), which were significant to the 2 canonical roots. *N*-heptacosane (C_{27}) occurs in higher concentrations in *M. consimilis*, and *n*-triacontane (C_{30}) occurs in higher concentrations in *M. cerberus* (see Table 2).



Figure 2. Dispersion diagram of discriminate analysis results showing the 2 canonical roots of differentiation based on the hydrocarbon profiles of *Mischocyttarus consimilis* and *M. cerberus* in colonies with and without interaction.

Table 2. Statistical values of the discriminate analysis of the most significant alkanes responsible for the distinction of *Mischocyttarus consimilis* and *M. cerberus* and of the individuals interacting parasitically, with average values of percentage area.

| HC | Nomenclature | Wilk's lambda | Р | Canonical coefficients | | % Relative area | | | |
|-------------------------------|---------------|---------------|--------|------------------------|--------|-------------------|-------------------|-------------------|--|
| | | | | Root 1 | Root 2 | M. consimilis | M. cerberus | On interaction | |
| C ₁₅ | n-pentadecane | 0.07 | < 0.00 | 1.36 | -0.09 | 0.11 ± 0.07 | 0.46 ± 0.04 | 0.25 ± 0.18 | |
| C ₁₆ | n-hexadecane | 0.04 | ns | | | | | | |
| C ₁₇ | n-heptadecane | 0.03 | ns | | | | | | |
| C_{18}^{17} | n-octadecane | 0.04 | ns | | | | | | |
| C ₁₉ | n-nonadecane | 0.04 | < 0.02 | 0.08 | -0.49 | 0.31 ± 0.25 | 0.25 ± 0.14 | 2.17 ± 2.64 | |
| C ₂₁ | n-heneicosane | 0.04 | ns | | | | | | |
| C_{23}^{21} | n-tricosane | 0.06 | < 0.00 | -0.31 | -0.52 | 16.59 ± 18.88 | 8.08 ± 15.29 | 19.96 ± 24.46 | |
| C24 | n-tetracosane | 0.04 | ns | | | | | | |
| C25 | n-pentacosane | 0.04 | ns | | | | | | |
| C_{26}^{23} | n-hexacosane | 0.05 | < 0.00 | 2.96 | -0.38 | 0.73 ± 1.17 | 3.37 ± 3.79 | 0.89 ± 1.36 | |
| C_{27}^{20} | n-heptacosane | 0.04 | < 0.01 | -0.24 | 0.74 | 18.23 ± 13.02 | 12.41 ± 9.40 | 6.28 ± 3.00 | |
| C_{28}^{27} | n-octacosane | 0.04 | ns | | | | | | |
| C_{29}^{28} | n-nonacosane | 0.05 | < 0.00 | -0.41 | -0.69 | 45.10 ± 22.15 | 39.13 ± 18.45 | 54.96 ± 22.98 | |
| C ₃₀ ²⁹ | n-triacontane | 0.06 | < 0.00 | 0.88 | 0.84 | 1.68 ± 1.16 | 13.42 ± 6.77 | 2.14 ± 1.08 | |

HC = hydrocarbons; ns = non-significant.

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Conversely, the chemical signatures of *M. consimilis* and *M. cerberus* are similar in colonies under interaction (see Figure 2). A greater overlap of statistical data of these interacting colonies was found compared to those of the 2 species without any interaction. A plausible conclusion is that the parasitic species can acquire part of the chemical signature of host colonies after first contact with their colonies, especially through contact with the substrate of their nests (Lorenzi and Cervo, 1992; Cervo and Lorenzi, 1996). Our results corroborate the conclusions of Cervo and Lorenzi (1996), who have observed that when females of *Polistes biglumis* invaded a host colony, they often vibrate their abdomens against the nest substrate, which suggests that the usurper species may attempt to acquire an odor similar to that of the host species.

The chemical signature of the host species changed after contact with the invader (see Figure 2), suggesting that the signatures of both species undergo modification through contact between individuals and nest structure.

The similarity of the chemical signatures of *M. cerberus* and *M. consimilis* in interactive colonies is defined by high concentrations of *n*-nonacosane (C_{29}) in the cuticles of both species (see Table 2). After the first contact between the 2 species, the concentration of *n*-nonacosane is altered. A similar phenomenon occurs in *P. biglumis*, in which the concentration of branched hydrocarbons in the nest material is altered owing to the overlap of the nest substrate, which is first realized by the queen of host colony and then by the usurper when this rubs its abdomen against the nest, intensifying the quantity of hydrocarbons present in the nest substrate (Lorenzi, 2011). Lorenzi and Cervo (1992) and Cervo and Lorenzi (1996) suggest that the female parasites acquire the profile of the host keeping contact with the material of the nest that seems to have similar elements to those present in the cuticle of individuals of the colony, which was noticed by Espelie and Hermann (1988, 1990), Espelie et al. (1990) and Singer et al. (1986a,b) found that contact with the nest substrate is important in *Polistes* species for the nestmates recognition via chemical signals.

Our results suggest that the 3 species of *Mischocyttarus* wasps studied have distinct chemical signatures, at least in terms of the linear hydrocarbons of their cuticles. However, these signatures are more alike in species with similar behavioral and morphological aspects. These similarities facilitate to occur a facultative parasitism interaction among the *M. consimilis* and *M. cerberus* species, in which the invader acquires part of the chemical signature of the host colony, which changes the chemical signature of both species after first contact, thus suggesting a chemical strategy to increase the acceptance of the invader by females of the host colony.

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